Synthesis of Tetracyclic Heterocompounds as Selective Estrogen Receptor Modulators. Part 3. Development of an Acid-Catalyzed Racemization Process for (*S*)-2,8-(Dimethoxy)-5-{4-[2-(1-piperidinyl)ethoxy]phenyl}-11,12-dihydro-*5H*-6,13-dioxabenzo[3,4]cyclohepta[1,2-*a*]naphthalene

Xun Li,*,[†] Ronald K. Russell,[†] András Horváth,[§] Nareshkumar Jain,[‡] Dominique Depré,[§] Dominic Ormerod,[§] Wim Aelterman,[§] and Zhihua Sui[‡]

Johnson & Johnson Pharmaceutical Research and Development, L.L.C., U.S. East Coast Research and Early Development, 1000 Route 202, Raritan, New Jersey 08869, U.S.A., and Johnson and Johnson Pharmaceutical Research and Development, A Division of Janssen Pharmaceutica, Turnhoutseweg 30, B-2340 Beerse, Belgium

Abstract:

A novel and economical process was developed for recycling the undesired enantiomer, (*S*)-2,8-dimethoxy-5-{4-[2-(1-piperidinyl)-ethoxy]phenyl}-11,12-dihydro-5*H*-6,13-dioxabenzo[3,4]cyclohepta[1,2-*a*]naphthalene (1b) obtained from chiral chromatographic separation, by refluxing 1b with HCl (4.0 equiv) in EtOH for 76 h, or with H₂SO₄ (2.0 equiv) in water for 68 h to afford a near racemic mixture ((*R*)-1a/(*S*)-1b = 41-42%/49-53%, chiral HPLC area%) in >96% isolated yield and good chemical purity (87-95%).

Introduction

The scale-up preparation of 2,5,8-substituted 11,12-dihydro-5H-6,13-dioxabenzo[3,4]cyclohepta-[1,2-a]naphthalene derivatives as selective estrogen receptor modulators (SERMs) has been reported recently,¹ where racemate **1** was prepared via an eight-step nonchromatographic linear synthetic process in 17% overall yield with 99.5% chemical purity (RPHPLC, area %). Chiral HPLC separation of 1 afforded (R)-enantiomer 1a and the corresponding (S)-enantiomer 1b (Scheme 1). Furthermore, the advanced in vitro and in vivo biological studies determined that (R)-1a was the most active compound with the desired SERM activity, while (S)-1b was the enantiomer with weak SERM activity. Because attempts for either enantioselective asymmetric synthesis of (R)-1a or the enantiomeric resolution of the racemate 1 with chiral acids were unfruitful,² the optically pure (>99% ee) (R)-1a was obtained on kilogram scale by the chiral chromatographic separation of racemate 1. The conversion of (S)-1b back to the racemic 1 would help in a higher yield of (R)-1a within a short time cycle. Herein, we report an acidcatalyzed racemization process for recycling (S)-1b.

Results and Discussion

A review of the literature showed no published results on the racemization of a tetracyclic compound like (R)-**1a**/(S)-**1b**,

[‡] Medicinal Chemistry Team VI, EC RED, U.S.A.

although it was known that the very similar (*R*)-3-(4-hydroxyphenyl)-4-methyl-2-(4-(2-(piperidin-1-yl)ethoxy)phenyl)-2*H*chromen-7-ol (**2a**, Scheme 2), a 2,3,4,7-substituted coumarin derivative, could be easily converted to a racemic mixture in 92% yield, after treatment with 5% LiOH in DMF at 80 °C for 3 h.³ Due to the similarity, compound (*R*)-**3a** (99.0% ee),⁴ the 2,8-dihydroxy analogue of (*R*)-**1a**, was first subjected to these known base-catalyzed conditions (5% LiOH in DMF at 80 °C) to produce a 63.3% (*R*)-**3a** enriched mixture after 60 h. The rate slowed unacceptably toward the end of the reaction, and no further optimization was conducted with LiOH-catalyzed racemization of (*R*)-**3a**.

When the above LiOH-catalyzed conditions were applied to pure 2,8-dimethoxy (*R*)-**1a** enantiomer (99.8% ee), however, no product (*S*)-**1b** was detected in the reaction mixture as analyzed by chiral HPLC. In addition, the treatment of (*R*)-**1a** separately with other bases (such as KOH, piperidine, and 4-(*N*,*N*-dimethyl)pyridine) in different refluxing solvents (such as MeOH, EtOH, and MeCN) also did not result in (*S*)-**1b**. It seemed that a base deprotonation of 2- and/or 8-phenolic hydroxyl group (calculated pK_{aS} of 2-OH and 8-OH = 10.19 and 9.88, respectively)¹ of (*R*)-**3a** was the driving force for this base-catalyzed racemization. The enantiomer (*R*)-**1a** was unable to be racemized under investigated basic conditions because of having 2,8-dimethoxy groups on the molecule.

The unsuccessful racemizations of (*R*)-**1a**/(*S*)-**1b** under basic conditions led us to explore other possible chemistries. For instance, (*R*)-**1a**/(*S*)-**1b** could possibly be racemized under acid-catalyzed conditions⁵ when more than one equivalent of an acid was used, due to the presence of the piperidine ring on the side

^{*} Author for correspondence. Telephone: (908)707-3321. Fax: (908)526-6469. E-mail: xli6@its.jnj.com.

[†] High Out Put Synthesis, EC RED, U.S.A.

[§] API Development, Janssen Pharmaceutica, Belgium.

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⁽⁴⁾ The optically pure (99.0% ee) (*R*)-enantiomer **3a** was obtained from a preparative chiral HPLC separation of its racemate **3**, which was prepared in-house and described as compound **11** in ref 1a of this publication.

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3a

Scheme 3

3a

chain of (*R*)-1a or (*S*)-1b. The excess acid could protonate the molecule on oxygen, preferably the 6-oxygen, to form a delocalized oxonium species, which might cause the C_5 stereogenic center to be racemized.^{5a}

To test this hypothesis, (S)-1b (98.6% ee) was treated with a slight excess (1.13 equiv) of either (1R)-(-)- or (1S)-(+)-10-camphorsulfonic acid ((-)-/(+)-CSA) (pK_a) $= -2.17)^{6a}$ in refluxing EtOH for 60 h; both reactions gave very similar (S)-1b-enriched ($\sim 68.5\%$, chiral HPLC area %) mixtures with the formation of (R)-1a ($\sim 29\%$) and a detectable level of byproduct **4b** ($\sim 2.3\%$)⁷ (Scheme 3, entries 1 and 2 of Table 1). Other solvents (such as CH₂Cl₂, MeCN, THF, and MeOH) were screened before EtOH was selected, which showed either no racemization (for example, in CH₂Cl₂ and MeCN) or less than 10% formation (in THF and MeOH) of (R)-1a, after (S)-1b (1.0 equiv) was refluxed with (1R)-CSA (1.13 equiv) in each tested solvent for 60 h, respectively. Among all acids tested, the best result (42.0% of (R)-1a) was achieved when only HCl (4.0 equiv, 1 M solution in EtOH) (pK_a = -6.0 and -6.1)^{6b,c} was used after 76 h (entry 8 of Table 1), extending the reaction time to 96 h did not change the product profile. Of interest, (R)-1a was also converted to a near racemic mixture with the presence of 4b (7.5%) under the above-described HCl-catalyzed conditions (entry 9 of Table 1). On the other hand, although the reaction of (S)-1b with 4.0 equiv of aqueous HCl (37% solution) in refluxing water for about three days that resulted in only trace amount (<1%) of (R)-1a (entry 13 of Table 1), H_2SO_4 (2.0 equiv) (p $K_a = -3.0$)^{6c} still accomplished a near racemic mixture under the same reaction conditions (entry 14 of Table 1). Considering HCl is a very strong Brønsted acid, other commonly used Lewis acids such as BF₃·Et₂O, MgBr₂·Et₂O, SnCl₄, and TiCl₄ were also examined for catalyzing the racemization of (S)-1b, all which, however, resulted in less than 10% of the desired enantiomer (R)-1a under the investigated conditions (entries 15-20 of Table 1). The above HCl-catalyzed reaction conditions were found to be reproducible on 1-, 10-, and 30-g scale of (S)-1b, which set the stage for racemization on larger scale. This acid-catalyzed process was not conducted on multikilogram scale because the project was unforeseeably terminated before the scale-up campaign.

3b

A proposed reaction mechanism for this acid-catalyzed racemization of (*S*)-**1b** is shown in Scheme 4. After the 6-oxygen of (*S*)-**1b** was protonated to the oxonium intermediate **5**, the positive charge could delocalize the electron density on C_5-O_6 bond via *path a* that led to the stabilized 4,5,8-trisubstitued-2,3-dihydrobenzo[*b*]oxepine carbocation intermediate **4a**,^{8a} of which the 4-substituted phenol hydroxyl group could approach to the sp² hybridized carbocation carbon^{8b} from either the top or bottom side and reclose the B-ring to achieve a racemic (*R*)-**1a**/(*S*)-**1b** mixture.⁵ On the other hand, the syn-

⁽⁷⁾ The structure of the byproduct 4b was confirmed by comparing its ¹H NMR, LC/MS (MH⁺ = 514), and HPLC analytical data with an inhouse prepared authentic compound 4b.

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Table 1. Results of acid-catalyzed racemization of (R)-1a and (S)-1b enantiomers with different acids in the selected solvents

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entry	cmpd	acid (equiv)	solvent	temp (°C)	time (h)	(<i>R</i>)-1a/(<i>S</i>)-1b/4b ^{<i>a</i>} (chiral HPLC, area %)
1	(S)- 1b (98.6% ee)	(1 <i>R</i>)-CSA (1.13)	EtOH	78	60	28.9/68.5/2.4
2	(S)-1b	(1 <i>S</i>)-CSA (1.13)	EtOH	78	60	28.8/68.7/2.3
3	(S)-1b	(1 <i>R</i>)-CSA (2.3)	EtOH	78	80	37.1/58.9/3.8
					150	39.7/57.5/2.8
4	(S)- 1b	TFA (2.3)	EtOH	70	60	5.3/94.5/0.28
5	(S)- 1b	MeSO ₃ H (4.0)	EtOH	75	68	38.3/61.7/ND ^e
6	(S)- 1b	$(1R)$ -CSA (1.0) + HCl $(3.0)^{b}$	EtOH	78	70	40.0/56.5/3.6
7	(S)- 1b	$(1R)$ -CSA (1.0) + HCl $(4.0)^{b}$	EtOH	78	60	41.1/53.8/5.1
8	(S)- 1b	HCl $(4.0)^{b}$	EtOH	78	76	42.0/53.1/4.8
9	(<i>R</i>)-1a (99.8% ee)	HCl $(4.0)^{b}$	EtOH	78	72	47.6/42.5/7.5
10	(S)- 1b	HCl $(4.0)^{c}$	EtOH	75	68	26.6/73.4/ND ^e
		HCl (20.0) ^c		75	68	42.1/57.9/ND
11	(S)- 1b	$MeSO_{3}H$ (4.0)	H ₂ O:EtOH (1:1)	75	68	24.5/75.5/ND
12	(S)- 1b	HCl $(4.0)^{c}$	H ₂ O:EtOH (1:1)	75	68	20.7/79.3/ND
		HCl (20.0) ^c		55	68	0.9/99.1/ND
13	(S)- 1b	HCl $(4.0)^{c}$	H_2O	95	68	0.8/99.2/ND
14	(S)- 1b	$H_2SO_4 (2.0)^d$	H_2O	95	68	41.0/49.0/2.5
15	(S)- 1b	$BF_3 \cdot Et_2O$ (1.0)	CH_2Cl_2	20	72	$0.5/99.5^{f}$
16	(S)- 1b	$BF_3 \cdot Et_2O$ (3.0)	THF	20	12	$0.5/99.5^{g}$
17	(S)- 1b	$MgBr_2 \cdot Et_2O$ (3.0)	CH_2Cl_2	40	12, 66	$7.5/92.5^h, 9.0/91.0^i$
18	(S)- 1b	SnCl ₄	CH_2Cl_2 (3.0)	20	12	$0.5/99.5^{j}$
19	(S)- 1b	$TiCl_4$ (1.0)	CH_2Cl_2	20	12	$3.5/96.5^k$
20	(S)- 1b	$TiCl_4$ (4.0)	THF	68	48	multiple products ¹

^{*a*} The retention times of the byproducts $4\mathbf{b} = 8.1$ min, (*R*)- $1\mathbf{a} = 11.4$ min, and (*S*)- $1\mathbf{b} = 22.3$ min as determined by chiral HPLC. ^{*b*} A 1.0 M solution in EtOH. ^{*c*} A 37% solution in H₂O. ^{*d*} A 96% solution. ^{*e*} Not determined. ^{*f*} Recovered 80% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*k*} Recovered 93% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*k*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*k*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} Recovered 91% of (*S*)- $1\mathbf{b}$ plus the rest of uncharacterized byproducts. ^{*i*} The mixture was uncharacterized.

Scheme 4. Proposed mechansism of acid-catalyzed racemization of (S)-enantiomer 1b



chronous elimination of an allylic proton from the 11-position of **5** via *path b* would result in the olefin byproduct **4b**.

Conclusions

The optically pure (*R*)-1a (99.8% ee) was unable to be converted to the racemic mixture 1 using basic conditions; this result was attributed to the absence of 2,8-phenolic protons on the molecule. In contrast, when the optically pure (*S*)-1b (98.6% ee) was treated with CSA in refluxing EtOH for 60–80 h, it produced an (*S*)-1b enriched mixture (~29%/69% to ~37%/ 59%; (*R*)-1a/(*S*)-1b). The addition of HCl to CSA further increased the yield of (*R*)-1a to greater than 40% of the mixture. The best reaction conditions were found with using HCl (4 equiv, 1.0 M in EtOH) to afford a near racemic mixture of (*R*)-1a/(*S*)-1b (42.0%/53.1%) in 99% isolated yield with high

chemical purity (>95%). When water was the solvent, 96% H_2SO_4 (2 equiv) also produced a near racemic mixture of (*R*)-1a/(*S*)-1b (41.0%/49.0%) in 96% isolated yield with good chemical purity (86.9%). Other Lewis acids displayed very weak (or no) catalytical effect to racemize (*S*)-1b. A new and economical process was developed for the reproducible recycling of the undesired (*S*)-1b isomer, which could be a useful method for the racemization of other enantiomerically pure 2,5,8-substituted 11,12-dihydro-5*H*-6,13-dioxabenzo[3,4]cyclohepta-[1,2-*a*]naphthalene derivatives whenever it is needed.

Experimental Section

The starting materials (*R*)-1a and (*S*)-1b were prepared inhouse, while the other reagents and solvents were obtained from commercial suppliers and were used without further purification.

¹H NMR spectra were recorded at 300 MHz on a Bruker Avance-300 instrument, and mass spectra were recorded on an Agilent series 180 LC/MS instrument (positive/negative modes). The chemical purity was determined on an Agilent series 1100 system at $UV_{max} = 254$ and 340 nm, using a ZORBAX Eclipse XDB-phenyl column (4.6 mm i.d. \times 50 mm, 3.5 micron) at 40 °C with flow rate of 1.0 mL/min and run time of 10.0 min. Solvent system: A - 80% H₂O + 0.1% TFA; B - 20% CH₃CN. Gradient: B 20% \rightarrow 80%/0.0 min \rightarrow 10.0 min, B 80% \rightarrow 90%/ 10.0 min → 11.0 min, B 90%/6.0 min, B 90% → 20%/17.0 $\min \rightarrow 20.0 \text{ min}$. The retention times of the byproduct 4b is 4.01 min and racemate 1 is 4.51 min. The optical purity/racemic ratio of (R)-1a/(S)-1b were determined on an Agilent series 1100 system at $UV_{max} = 210$ and 254 nm, using a Chiralpak AD column (Chiral Technologies, Inc.) (4.6 mm ID × 250 mm, 10 micron) at 20 °C with flow rate of 1.0 mL/min and run time of 33.0 min with 100% 2-propanol as the mobile phase. The retention time of the byproduct 4b is 8.1 min, (R)-1a is 11.4 min, and (S)-1b is 22.3 min. Preparative chiral HPLC separation of (R)-1a/(S)-1b was conducted on a Chiralpak AD column $(5.0 \text{ cm i.d.} \times 150 \text{ mm}, 10 \,\mu\text{m})$, at 20 °C with flow rate of 50 mL/min and run time of 25.0 min, with 100% 2-propanol as the mobile phase.

A Typical Acid-Catalyzed Racemization Procedure in EtOH for (S)-Enantiomer 1b. A four-neck 500-mL roundbottom flask (RBF) equipped with a thermocouple controller, a mechanical stirrer, a condenser, a pressure-equalization dropping funnel, a septum, and a nitrogen inlet adapter was charged with (S)-1b (10.0 g, 0.0195 mol; 98.6% ee) and EtOH (120.0 mL, 190 proof), and the suspension was stirred under nitrogen. A solution of HCl (77.8 mL, 0.0778 mol; 1.0 M in EtOH) was added over a 10-min period (this addition was a mildly exothermic process, the internal temperature was 28 °C after the addition), and the reaction was heated to reflux (78 °C, internal temperature) for 76 h. The progress of the racemization was determined by chiral HPLC analysis (authentic racemate 1 (>99.5%) was used as a standard reference). After the reaction time, the reaction was cooled to 20 °C, and the solvent was concentrated at 60 °C under house vacuum (~120 mmHg). The resulting nearly racemic HCl salt was a dark-cherry, foamy solid, which was dissolved in D.I. H₂O (100 mL) and cooled to 0 °C in an ice-water bath, and the pH of the solution was adjusted to ≥ 12.0 with 5 N NaOH solution (~40 mL). The alkaline solution was extracted with EtOAc (150 mL \times 3), and the combined organic phase was washed with brine (100 mL). The solvent was concentrated at 60 °C under house vacuum to afford 10.56 g (105.6% isolated yield) of the near-racemic 1 free base as a yellow-brown, foamy solid. The identity and

purity of this material was confirmed by comparing the analytical data of its ¹H NMR, LC/MS, the retention times on reverse phase HPLC (97%, area %) and chiral HPLC (>95%, total area %) with that of an in-house prepared authentic racemate **1**. ¹H NMR (300 MHz, CDCl₃) δ 1.41 (m, 2 H), 1.58 (m, 4 H), 2.46 (m, 4 H), 2.71 (t, *J* = 6.0, 2 H), 2.89 (t, *J* = 5.8, 2 H), 3.73 (s, 3 H), 3.79 (s, 3 H), 4.03 (t, *J* = 6.1, 2 H), 4.69 (t, *J* = 5.5, 2 H), 6.06 (s, 1 H), 6.36 (d, *J* = 1.0, 2 H), 6.48 (dd, *J* = 0.9, 8.3, 1 H), 6.58 (dd, *J* = 0.9, 8.2, 1 H), 6.68 (d, *J* = 9.1, 2 H), 7.02 (d, *J* = 8.4, 1 H), 7.17 (d, *J* = 8.3, 1 H), 7.36 (d, *J* = 9.0, 2 H). LC/MS *m*/*z* 514 (MH⁺), 536 (MNa⁺).

A Typical Acid-Catalyzed Racemization Procedure in H₂O for (S)-Enantiomer 1b. Into a four-neck 500 mL RBF were charged (S)-1b (10.8 g, 0.021 mol, 93.2%) and water (250 mL). After the yellowish suspension was warmed to 60 °C, a solution of 96% H₂SO₄ (2.24 mL, 0.042 mol) was added dropwise, and the formed yellow-orange suspension was heated further to 95 °C (which became a clear, yellowish solution at 80 °C) and stirred for 68 h. After the reaction was cooled to 80 °C, toluene (100 mL) was added to the brownish mixture, and then the pH was adjusted to ~ 10 by addition of 50% aqueous NaOH (4 mL) with intensive stirring. The resulting mixture was cooled to 50 °C, diluted with THF (50 mL), and further cooled to 20 °C with intensive stirring. The two phases were then separated, and the organic layer was evaporated to afford 10.4 g (96.3% isolated yield; 87%, HPLC area %) of the nearracemic 1 free base as a sticky, brown, foamy solid.

Chiral chromatographic purification of the above obtained near-racemic **1** free base (8.0 g) afforded: 3.31 g (41% recovery yield; 95.9%, RPHPLC, area %; 99.9% ee, chiral HPLC) of the desired (R)-**1a** containing 1.55% (RPHPLC, area %) of **4b** and the rest of uncharacterized byproduct; plus 3.92 g (49% recovery yield; 96.6%, RPHPLC, area %; 99.2% ee, chiral HPLC) of the undesired (S)-**1b**, which also contained 0.95% (RPHPLC, area %) of **4b**.

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